

# Immunoprecipitation/IP Kit-Anti-CALML3 Immunomagnetic Beads

Catalog Number: MB12480-RP02

Please read this instruction manual carefully before using the product

#### **Product Contents**

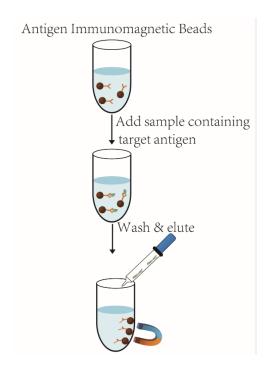
Contents	Package 1	Package 2	Storage
Anti-CALML3 Immunomagnetic Beads <sup>13</sup>	1 mL	5 mL	2-8°C for 12 months
NP40 Cell Lysis Buffer	4 mL	22 mL	-20°C for 12 months
5×TBST (pH7.4)	Required but not supplied		
1×TBST (pH7.4)	Required but not supplied		
$ddH_2O$	Required but not supplied		
Alkaline Elution Buffer	3 mL	15 mL	2-8℃ for 12 months
Acidity Elution Buffer	3 mL	15 mL	2-8℃ for 12 months
Neutralization Buffer	2 mL	8 mL	2-8℃ for 12 months
Magnetic Separator	Not included (refer related product MAGS001)	One MAGS001 included in China <sup>2</sup>	

- [1] The IP KIT contains anti-CALML3 Immunomagnetic Beads(2 mg/mL) in phosphate buffered saline (PBS, pH 7.4) with sodium azide (0.1%).
- [2] The Magnetic Separator cannot be included for oversea customers due to shipment prohibition.
- [3] Immunomagnetic Beads kits are shipped at ambient temperature in which immunomagnetic beads are provided in liquid buffer.

## **Product Description**

The Anti-CALML3 Immunomagnetic Beads, conjugated with Anti-CALML3 antibody, are used for immuneprecipitation (IP) of CALML3 proteins which expressed in vitro expression systems and bacterial and mammalian cell lysates.

For IP, the beads are added to a sample containing CALML3 proteins to form a bead-protein complex. The complex is removed from the solution manually using a Magnetic Separator. The bound CALML3 proteins are dissociated from the Immunomagnetic Beads using an elution buffer.



# **Antibody Information**

Antibody: CALML3 Antibody, Rabbit PAb, Antigen Affinity

Purified(12480-RP02)

Immunogen: Recombinant Human CALML3 protein

(Catalog#12480-H07E)

Clone ID:

Isotype: Rabbit IgG

Specificity: Human CALML3

**Guaranteed Applications:** IP, Minimum Protein Purification **Preparation:** Produced in rabbits immunized with purified, recombinant Human CALML3 (rh CALML3; Catalog#12480-H07E; P27482; Met1-Lys149). CALML3 specific IgG was purified by

Human CALML3 affinity chromatography.

Fig. 1 Immunoprecipitation (IP) Protocol

#### **Protocol**

The protocol (Fig. 1) uses  $50 \mu L$  Anti-CALML3Immunomagnetic Beads, but this can be scaled up or down as required.

#### **Cell Lysis**

Cells may be lysed using any standard cell lysis protocol in accordance with your starting materials. We suggest using NP40 Cell Lysis Buffer (supplied with kit).

#### **Immunoprecipitate Target Antigen**

- 1. Add 50  $\mu$ L of Immunomagnetic Beads into a 1.5 mL microcentrifuge tube.
- 2. Add 150  $\mu$ L of 1 $\times$  TBST buffer to the Immunomagnetic Beads and gently vortex to mix.
- 3. Place the tube into a Magnetic Separator to collect the beads against the wall side of the tube. Remove and discard the supernatant.
- 4. Add 1 mL of 1×TBST buffer to the tube. Invert the tube several times or gently vortex to mix for 1 min. Collect Immunomagnetic Beads with a Magnetic Separator. Remove and discard the supernatant.
- 5. Add the sample containing target protein (~100  $\mu$ g of protein in 100  $\mu$ L) to the pre-washed Immunomagnetic Beads, add 400  $\mu$ L of 1 $\times$ TBST buffer and incubate at room temperature for 30 min with mixing.
- Collect the Immunomagnetic Beads with a Magnetic Separator, remove the unbounded sample and save for analysis.
- 7. Add 300  $\mu$ L of 5  $\times$  TBST buffer to the tube and gently mix. Collect the Immunomagnetic Beads and discard the supernatant. Repeat this wash twice.
- 8. Add 300  $\mu$ L of ddH<sub>2</sub>O to the tube and gently mix. Collect the Immunomagnetic Beads on a Magnetic Separator and discard the supernatant.

#### Elute Target Antigen.

- A. Alkaline Elution Protocols
- 1. Add 100  $\mu$ L of Alkaline Elution buffer to the tube.
- 2. Gently vortex to mix and incubate the sample at room temperature on a rotator for 5 min.
- 3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the target antigen.
- 4. To neutralize the sample, add 50  $\mu L$  of Neutralization Buffer for each 100  $\mu L$  of eluate.
- **B.** Acidity Elution
- 1. Add 100 μL Acidity Elution Buffer.
- 2. Gently vortex to mix and incubate the sample at room temperature on a rotator for 5-10 min.
- 3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the target antigen.

- 4. To neutralize the low pH, add 15  $\mu L$  of Neutralization Buffer for each 100  $\mu L$  of eluate.
- C. Elution Using Sample Buffer
- 1. Add 100  $\mu$ L of SDS-PAGE sample buffer to the tube.
- 2. Gently vortex to mix and incubate the sample at  $95-100^{\circ}$ C for 5-10 min.
- 3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the antigen.

# **Reference Information**

### **Related Products**

Products	Cat No.
Magnetic Separator-1.5 (2 tubes)	MAGS001
Immunoprecipitation Kit -Immunomagnetic Beads Protein A Kit	BA10600
Immunoprecipitation Kit -Immunomagnetic Beads Protein G Kit	BG13103
Immunoprecipitation Kit -Immunomagnetic Beads Protein L Kit	BL11044
Immunoprecipitation Kit -Anti-DYKDDDDK(Flag®) Tag Immunomagnetic Beads Kit	TB101274
Immunoprecipitation Kit -Anti-GFP Tag Immunomagnetic Beads Kit	TB13105
Immunoprecipitation Kit -Anti-Myc Tag Immunomagnetic Beads Kit	TB100029
Immunoprecipitation Kit -Anti-HA Tag Immunomagnetic Beads Kit	TB100028
Immunoprecipitation Kit -Anti-V5 Tag Immunomagnetic Beads Kit	TB100378

## **Trouble Shooting**

Problem	Possible Cause	Solution
Little or no protein is detected	Protein degraded	Include protease inhibitors (PMSF) in the lysis buffer
		Use new lysate or lysate stored at -80° C
	No or minimal protein was expressed	Verify protein expression by SDS-PAGE or Western blot
		Analysis of the lysate using an positive control as a reference
		Increase the amount of lysate used for IP/Co-IP
		Use a more sensitive detection system

Problem	Possible Cause	Solution	
Magnetic Beads aggregated	Magnetic Beads were frozen or centrifuged	Handle the Beads as directed in the instructions	
	Buffer was incompatible with magnetic beads		
	Detergent was not added to the wash and bind solutions		
Failure to co-IP interacting protein	Wash conditions were too stringent	Reduce the number of washes	
	for the weak or transient interaction	Lower the ionic strength of the wash buffer	
	Interacting protein was expressed at a low level	Apply additional protein sample	
		Use a more sensitive detection system	
	Buffer system was not optimal for the protein: protein interaction	Optimize the co-IP buffer	
	Insufficient	Elute sample in 30% acetonitrile 0.5% formic acid, then	
	sample was loaded on the gel for Western blot detection	Bring the sample back up in SDS-PAGE Sample Buffer and load entire elution fraction on	