Designation: TF-1

Cryovial: 300434 Vital: 330434 CLS order number:



Origin and General Ch	aracteristics
Organism:	Homo sapiens (human)
Ethnicity:	Japanese
Age:	35 years of age
Gender:	Male
Tissue:	Bone marrow
Morphology:	Lymphoblast
Cell type:	Erythroleukemia; erythroblast
Growth Properties:	Suspension
Description:	The TF-1 cell line has been established by T. Kitamura in October 1987 from a heparinised bone marrow aspiration sample from an erytholeukemic patient. TF-1 cells proliferate depending on the GM-CSF and IL-3. The morphological and cytochemical features, plus the constitutive expression of globin genes, indicate the commitment of the cells to the erythroid lineage. TPA induces a dramatic differentiation into macrophage-like cells.
References:	Kitamura T et al. Establishment and characterization of a unique human cell line that proliferates dependently on GM-CSF, IL-3, or erythropoietin. J Cell Physiol 140: 323-34, 1989.
Culture Conditions and	Handling
Culture Medium:	RPMI 1640 medium supplemented with 4.5g/L glucose, L-glutamine,10% KMG, and 10% fetal bovine serum (MG-75, CLS order number 820705). [for long term culture, TF-1 cells need interleukin 3 (IL-3, GM-CSF) in the culture medium].
Subculturing:	Start cultures at 2 x 10 <sup>5</sup> cells/ml and maintain between 2 x 10 <sup>5</sup> and 1 x 10 <sup>6</sup> cells/ml. Culture at 37°C/5% CO <sub>2</sub> .  Subculture by transferring an aliquot of the cell suspension into a new cell culture flask already containing an appropriate amount of fresh cell culture medium.
Split Ratio:	every 2 to 3 days
Seeding density:	Minimum of 2 x 10 <sup>5</sup> cells/ml
Fluid Renewal:	Feed the cells by collecting the cells by centrifugation, remove the supernatant and add the same volume of fresh cell culture media to the cell pellet. Resuspend and transfer into the cell culture flask.
Freeze Medium:	CM-1 (CLS order number: 800125, 25ml, 800150, 50ml)
Freezing recovery:	every 2 to 3 days
Sterility:	Mycoplasma specific PCR: negative; Bacteria specific PCR: negative
Biosafety Level:	1
Safety precautions:	If the cryovial is planned to be stored in liquid nitrogen and to be thawed in the future, special safety precautions should be followed:  Protective gloves and clothing should be used and a facemask or safety goggles must be worn when transferring frozen samples into or removing from the liquid nitrogen tank.  The removal of a cryovial from liquid nitrogen may result in the explosion of the frozen vial creating flying fragments.  Caputo, J.L. Biosafety procedures in cell culture. J. Tissue Cult. Methods 11:223-227, 1988. ATCC Quality Control Methods for Cell Lines, 2nd edition, 1992.
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	fetal bovine serum (MG-75, CLS order number 820705). [for long term culture, TF-1 cells need interleukin 3 (IL-3, GM-CSF) in the culture medium].	
Special Features of the	Cell Line	
Receptors Expressed:	TF-1 cells do not express glycophorin A or carbonyl anhydrase I.	
DNA Profile (STR):	Amelogenin: X,Y	vWA: 15,17
	CSF1PO: 13 D13S317: 8,9	D3S1358: 15 D21S11: 30
	D16S539: 9,12 D5S818: 13	D18S51: 13 Penta E: 5,17
	D7S820: 12 TH01: 7,9	Penta D: 10,13 D8S1179: 11,15
	TPOX: 8	FGA: 18,19
Applications:	The TF-1 cell line can be applied in various systems due to their responsiveness to multiple cytokines. They provide a good system to investigate the proliferation and differentiation of myeloid progenitor cells.	

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	Certificate of Analysis:	The Certificate of Analysis for each batch can be requested by e-mail at	
		service@clsgmbh.de.	

Recommendations for handling of cells growing in suspension following delivery	
Cryopreserved cells	The cells come deep-frozen shipped on dry ice. Please make sure that the vial is still frozen.
	If immediate culturing is not intended, the cryovial(s) must be stored below -150°C after arrival.
	If immediate culturing is intended, please follow these instructions:
	Quickly thaw by rapid agitation in a 37°C water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. Note: A small ice clump should still remain and the vial should still be cold.
	From now on, all operations should be carried out under aseptic conditions.
	Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300xg for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later.
	Resuspend the cells carefully in 10ml fresh cell culture medium and transfer them into one T25 cell culture flask. All further steps are described in the Subculture section.
Proliferating Cultures	The cell culture flask, 1xT25, comes filled with cell culture medium.
	Incubate at 37°C for a minimum of 24 hrs.
	Count the cells, spin down the cell suspension at 300x g for 3 minutes to collect the cells. Resuspend the cells in an appropriate amount of fresh cell culture medium and transfer to new cell culture flasks.
	Incubate at 37°C for a minimum of 24 hrs.

Warranty:	CLS warrants for a high cell viability and culture performance only if the product(s) is (are) stored and cultured according to the information described above. Using cell culture media and supplements other than the ones recommended in this product information may result in satisfactory proliferation and viabilities. CLS, however, does not warrant for cell recovery, proliferation and function if differing formulations are employed.
Disclaimer:	The customer shall not be entitled to employ this product for purposes other than research. Commercial utilization shall not be permitted; in particular, the cell line, its components or materials made therefrom shall not be sold or transferred to any third party. In addition, the term 'Commercial use' shall mean any activity by a party for consideration and may include, but is not limited to, use of the product or its components

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