



## Fluorescent Green qPCR Master Mix (high cxr) (250 rxns)

Zellbio GmbH (Germany)

CAT No. ZX-22113-250

[www.zellx.de](http://www.zellx.de)

Post reverse transcription step detection and quantification of DNA and cDNA targets, low copy gene detection, Gene expression using standard and fast qPCR platforms

**!!! Caution: This product is for Research Use Only. Not for *in-vitro* Diagnostics !!!**

## Table of Contents

<b>Introduction</b> .....	3
<b>Materials supplied in the Kit</b> .....	3
<b>Storage instruction</b> .....	3
<b>Materials required but not supplied</b> .....	3
<b>Precautions</b> .....	3
<b>General remarks</b> .....	3
<b>Assay Procedure</b> .....	4

Please read this insert completely prior to using the product.

## Introduction

ZellX® Fluorescent Green qPCR Master Mix (high cxr) grants efficient quantitative/real-time PCR in a single tube. The Fluorescent Green qPCR Master Mix contains a green fluorescent reporter dye plus all the reagents (except primers and DNA template) needed for running real-time PCR reactions. As an internal reference, the kit contains high concentrations of carboxy-X-rhodamine (ROX™). Being independent from the amount of DNA template, the fluorescence signal of ROX™ is not influenced by the PCR reactions, and therefore can assist in the normalization of the reporter-dye signal during data analysis. The appropriate level of ROX™ depends on the real-time PCR instrument (Contact your instrument manufacturer for details). For low levels of ROX™ use our **Fluorescent Green qPCR Master Mix (low cxr) (Cat NO. ZX-22112-250/500/1000)**.

## Materials supplied in the Kit

<i>Component</i>	<i>Quantity</i>
qPCR Master Mix (2X)	2 x 1.25 mL

## Storage instruction

All reagents should be stored at -20°C upon receipt. Avoid repeated freezing and thawing.

## Materials required but not supplied

Precision pipettes and disposable filter pipette tips (RNase & DNase free)

Nuclease-free tubes / strips / plates corresponding to the PCR device

## Precautions

This kit should only be used by qualified personnel who have had laboratory safety instruction. The complete insert should be read and understood before attempting to use the product.

## General remarks

- The instruction must be strictly followed. PCR machine / Thermocycler must be turned on and programmed in advance to avoid delays after setting up the reactions.
- Pipette tips should not be used more than once to prevent cross contamination.
- Reagents of different batches should not be mixed or used after their expiration dates.

## Assay Procedure

### For 20 $\mu$ L reaction

- Thaw Master Mix on ice and mix them well. Collect liquid at the bottom of the tube with a quick spin.
- Set up the following reaction mixture.

<b>Component</b>	<b>Quantity</b>
<b>qPCR Master Mix</b>	10 $\mu$ L
<b>Forward Primer</b>	X $\mu$ L (200 nM)
<b>Reverse Primer</b>	X $\mu$ L (200 nM)
<b>DNA template</b>	10-100 ng
<b>Nuclease-Free Water</b>	Up to 20 $\mu$ L

*\*For optimal performance, we recommend to use cDNA corresponding to 1 pg to 500 ng of total RNA. For genomic DNA, we recommend to not use more than 100 ng*

- Mix reagents thoroughly, and transfer to the thermocycler.
- Run the appropriate PCR cycling protocol on your real-time PCR instrument

<b>Step</b>	<b>Number of Cycles</b>	<b>Temperature</b>	<b>Duration</b>
<b>Initial activation</b>	1	95°C	30 sec
<b>Amplification*</b>	40	95°C	3-5 sec
		60-65°C*	20-30 sec

*\*Not < 60°C.*

- The appropriate PCR cycling protocol must be optimized by the end user
- high primer concentrations result in nonspecific amplification and should be avoided