

# Monkey IgM ELISA

## Life Diagnostics, Inc., Catalog Number: IGM-3-INT

### Monkey IgM ELISA (For Export)

#### INTRODUCTION

The monkey IgM ELISA kit is designed for measurement of IgM in old world monkey serum or plasma. The assay uses a mouse monoclonal anti-monkey IgM antibody (LDI, clone 2C11-1-5) for solid phase (microtiter wells) immobilization. The same antibody, conjugated to horseradish peroxidase (HRP), is used for detection. The assay detects IgM from species including Baboon, Rhesus, Cynomolgus and Squirrel monkeys.

#### PRINCIPLE OF THE TEST

Test samples are diluted and incubated in the microtiter wells for 45 minutes alongside prepared IgM standards. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. IgM molecules are thus sandwiched between the immobilization and detection antibodies. The wells are then washed to remove unbound HRP conjugate and TMB Reagent is added and incubated for 20 minutes. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow. Optical density is measured spectrophotometrically at 450 nm. The concentration of IgM is proportional to the optical density of the test sample and is derived from a standard curve.

#### MATERIALS AND COMPONENTS

##### Materials provided with the kit:

- Anti monkey IgM coated 96-well plate (12 strips of 8 wells)
- HRP Conjugate, 11 ml
- Reference standard (lyophilized)<sup>1</sup>
- 20x Wash Solution: TBS50-20, 50 ml
- 10x Immunoglobulin Diluent: RD25-10, 25 ml
- TMB Reagent (One-Step): TMB11-1, 11 ml
- Stop Solution (1N HCl): SS11-1, 11 ml

##### Materials required but not provided:

- Precision pipettes and tips
- Distilled or deionized water
- Polypropylene or glass tubes
- Vortex mixer
- Absorbent paper or paper towels
- Micro-Plate incubator/shaker mixing speed of  $\approx 150$  rpm
- Plate washer
- Plate reader with an optical density range of 0-4 at 450 nm
- Graph paper or PC graphing software

#### STORAGE OF THE TEST KIT

The test kit will remain stable for six months from the date of purchase provided that the components are stored at 2-8°C. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air.

<sup>1</sup> The IgM standard used in this kit is of human origin. It behaves identically to Rhesus and Cynomolgus monkey IgM. The use of a human standard allows export of the kit without requirement for CITES documentation.

#### GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (25°C) before use.
3. Optimum results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

#### DILUENT PREPARATION

The diluent is provided as a 10x stock. Prior to use, estimate the final volume of diluent required for your assay and dilute one (1) volume of the 10x stock with nine (9) volumes of distilled or deionized water.

#### WASH SOLUTION PREPARATION

The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.

#### STANDARD PREPARATION

1. The IgM standard is provided as a lyophilized stock. Reconstitute with 1.0 ml of distilled or deionized water (***the reconstituted standard is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended***).
2. Label 7 polypropylene or glass tubes as 250, 125, 62.5, 31.25, 15.63, 7.81, and 3.91 ng/ml.
3. Into the tube labeled 250 ng/ml, pipette the volume of diluent detailed on the IgM standard vial label. Then add the indicated volume of IgM standard (shown on the IgM standard vial label) and mix gently. This provides the 250 ng/ml standard.
4. Dispense 250  $\mu$ l of diluent into the tubes labeled 125, 62.5, 31.25, 15.63, 7.81, and 3.91 ng/ml.
5. Prepare a 125 ng/ml standard by diluting and mixing 250  $\mu$ l of the 250 ng/ml standard with 250  $\mu$ l of diluent in the tube labeled 125 ng/ml.
6. Similarly prepare the 62.5, 31.25, 15.63, 7.81, and 3.91 ng/ml standards by serial dilution.

#### SAMPLE PREPARATION

**General Note:** IgM is typically present in monkey serum or plasma at concentrations of  $\approx 0.3$  mg/ml. In order to obtain values within range of the standard curve, we suggest that samples initially be diluted 10,000-fold using the following procedure for each sample to be tested:

1. Dispense 247.5  $\mu$ l of 1x diluent into two tubes.
  2. Pipette and mix 2.5  $\mu$ l of the serum/plasma sample with 247.5  $\mu$ l of 1x diluent in the first tube. This provides a 100-fold diluted sample.
  3. Mix 2.5  $\mu$ l of the 100-fold diluted sample with the 247.5  $\mu$ l of diluent in the second tube. This provides a 10,000-fold dilution of the sample.
  4. Repeat this procedure for each sample to be tested.
- Tissue extracts and body fluids other than serum or plasma will likely have lower IgM levels than those found in serum. Optimal dilutions of such samples should be determined empirically.

### ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100  $\mu$ l of standards and diluted samples into the wells (we recommend that samples and standards be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at 25°C for 45 minutes<sup>2</sup>.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400  $\mu$ l/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100  $\mu$ l of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at 25°C for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100  $\mu$ l of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at 25°C for 20 minutes.
11. Stop the reaction by adding 100  $\mu$ l of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader *within 5 minutes*.

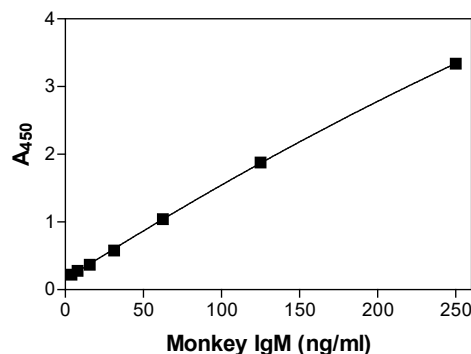
### CALCULATION OF RESULTS

1. Calculate the average absorbance values ( $A_{450}$ ) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in ng/ml on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of IgM in ng/ml from the standard curve.
4. Multiply the derived concentration by the dilution factor to determine the actual concentration of IgM in the sample.
5. PC graphing software may be used for the above steps.
6. If the OD<sub>450</sub> values of samples fall outside the standard curve, samples should be diluted appropriately and re-tested.

### TYPICAL STANDARD CURVE

A typical standard curve with optical density readings at 450nm on the Y-axis against IgM concentrations on the X-axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

IgM (ng/ml)	Absorbance (450 nm)
250	3.34
125	1.879
62.5	1.041
31.25	0.579
15.63	0.368
7.81	0.278
3.91	0.222



### LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings

Rev 02202020

For technical assistance please email us at [techsupport@lifediagnosics.com](mailto:techsupport@lifediagnosics.com)

<sup>2</sup> The kit was validated using a shaking incubator at 25°C and 150rpm. Performance of the assay at lower temperature and/or lower temperatures will result in lower absorbance values.