

5-Hydroxyindoleacetic Acid (5-HIAA) ELISA Kit**Catalog No.:** abx257174**Size:** 96T**Range:** 1.563 - 100 ng/ml**Sensitivity:** 0.938 ng/ml**Storage:** Store at 4°C for up to 6 months

Application: For quantitative detection of 5-Hydroxyindoleacetic Acid (5-HIAA) in serum, plasma, tissue homogenates and other biological fluids.

Introduction

5-HIAA is the main metabolite of serotonin. In chemical analysis of urine samples, 5-HIAA is used to determine serotonin levels in the body. 5-HIAA levels can vary depending on other complications, including tumors, renal malfunction, and small bowel resection.

Principle of the Assay

This kit is based on a competitive enzyme-linked immuno-sorbent assay technology. 5-HIAA is pre-coated onto 96-well plates. The standards, samples and a biotin conjugated antibody specific to 5-HIAA are added to the wells and incubated. After washing away the unbound conjugates, HRP Streptavidin (SABC) is added to each microplate well and incubated. After TMB substrate solution is added only wells that contain 5-HIAA will produce a blue colour product that changes into yellow after adding stop solution. The density of yellow is proportional to the 5-HIAA amount of sample captured in plate. The O.D. absorbance is measured spectrophotometrically at 450nm in a microplate reader, and then the concentration of 5-HIAA can be calculated.

Kit Components

1. One pre-coated 96 well plate
2. Standards: 2 tubes
3. Sample/Standard diluent buffer: 20 ml
4. Biotin conjugated antibody (Concentrated): 60 µl, Dilution 1:100
5. Antibody diluent buffer: 10 ml
6. HRP streptavidin conjugate (SABC) (Concentrated): 120 µl, Dilution 1:100
7. SABC diluent buffer: 10 ml
8. TMB substrate: 10 ml
9. Stop solution: 10 ml
10. Wash buffer (25X): 30 ml

Material Required But Not Provided

1. 37°C incubator
2. Microplate reader (wavelength: 450nm)
3. Precision pipette and disposable pipette tips
4. Automated plate washer
5. ELISA shaker
6. 1.5ml tubes to prepare standard/sample dilutions
7. Plate cover
8. Absorbent filter papers
9. 100 ml and 1 L volume graduated cylinder.

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Protocol

A. Preparation of sample and reagents

1. Sample

Store samples to be assayed within 24 hours at 2-8°C. Alternatively, aliquot and store at -20°C or -80°C for long term. Avoid repeated freeze-thaw cycles.

- ✧ **Tissue homogenates:** The preparation of tissue homogenates will vary depending upon tissue type – this is just an example. Rinse tissues with ice-cold PBS (0.01 mol/L, pH 7.2) to remove the excess of blood. Weigh before homogenization. Finely mince tissues and homogenize with a tissue homogenizer on ice in PBS and sonicate the cell suspension. Centrifuge the homogenates at 5000 × g for 5 min and collect the supernatant. Assay immediately or aliquot and store at -20°C.
- ✧ **Serum:** Allow the serum to clot in a serum separator tube at room temperature (2 hours). Centrifuge at approximately 1000 × g for 20 minutes. Analyze the supernatant immediately or aliquot and store at -20°C.
- ✧ **Plasma** Collect plasma using EDTA-Na₂ as an anticoagulant. Centrifuge for 15 minutes at 1000 × g within 30 minutes of collection. Assay immediately or aliquot and store at -20°C. Avoid hemolysis and high cholesterol samples.
- ✧ **Other biological fluids:** Centrifuge at approximately 1000 × g for 20 min to remove precipitant. Analyze immediately or aliquot and store at -20°C or -80°C.

Note:

- Fresh samples or recently obtained samples are recommended to prevent degradation and denaturalization that may lead to erroneous results.
- Coagulate blood samples completely, centrifuge, and avoid hemolysis and precipitant. Hemolysis will influence the result. Please bring samples slowly to room temperature.

2. Wash buffer

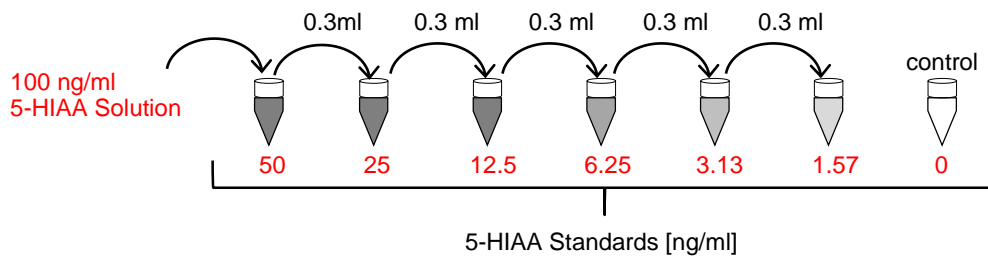
Dilute the concentrated Wash buffer 25-fold (1/25) with distilled water (i.e. add 30 ml of concentrated wash buffer into 720 ml of distilled water).

3. Standard

Preparation of the 5-HIAA standard: standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of standard are included in each kit. Use one tube for each experiment. **(Note: Do not dilute the standard directly in the plate).**

a.) 100 ng/ml standard solution. Add **1 ml** of Sample/Standard diluent buffer into one Standard tube, keep the tube at room temperature for 10 min, mix thoroughly and avoid foaming or bubbles.

b.) 50 ng/ml → 1.57 ng/ml standard solutions: Label 6 tubes with 50 ng/ml, 25 ng/ml, 12.5 ng/ml, 6.25 ng/ml, 3.13 ng/ml and 1.57 ng/ml. Aliquot **0.3 ml** of the Sample / Standard diluent buffer into each tube. Add **0.3 ml** of the above 100 ng/ml standard solution into 1st tube and mix thoroughly. Transfer **0.3 ml** from 1st tube to 2nd tube and mix thoroughly. Transfer **0.3 ml** from 2nd tube to 3rd tube and mix thoroughly, and so on.



Note: The standard solutions are best used within 2 hours. The standard solution can be stored at 4°C for up to 12 hours, or at -20 °C for up to 48 hours. Avoid repeated freeze-thaw cycles.

4. Preparation of Biotin conjugated antibody working solution: prepare no more than 1 hour before the experiment.

a.) Calculate the total volume of the working solution: $0.05 \text{ ml / well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).

b.) Dilute the Biotin conjugated antibody with Antibody diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of Biotin conjugated antibody into 99 μl of Antibody diluent buffer.

5. Preparation of HRP Streptavidin Conjugate (SABC) working solution: prepare no more than 1 hour before the experiment.

a.) Calculate the total volume of the working solution: $0.1 \text{ ml / well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).

b.) Dilute the SABC with SABC diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of SABC into 99 μl of SABC diluent buffer.

B. Assay procedure

Equilibrate the SABC working solution and TMB substrate for at least 30 minutes to room temperature prior to use. It is recommended to plot a standard curve for each test.

1. **Wash the plate two times before adding standard, samples and buffers.** Any strips that are not being used should be kept dry and stored at 4°C. Set standard, test sample and control (zero) wells on the pre-coated plate and record their positions. It is recommended to measure each standard and sample in duplicate.
2. Add 50 μl of the prepared standards solutions into the standard wells.
3. Add 50 μl of Sample / Standard diluent buffer into the control (zero) well.
4. Add 50 μl of appropriately diluted sample into test sample wells.
5. Immediately add 50 μl of Biotin conjugated antibody working solution into each well. (Please add the solution at the bottom of each well without touching the side wall).
6. Cover the plate with the plate sealer. Gently tap the plate to mix thoroughly. Incubate at 37°C for 45 minutes.
7. Remove the cover, and wash the plate 3 times. Discard the solution without touching the side walls. Blot the plate on an absorbent material. Fill each well completely with wash buffer and soak for at least 1-2 min. Discard the contents and clap the plate on absorbent filter papers or other absorbent material. Repeat this procedure for a total of three times.

Please note: For automated washing, discard the solution in all wells and wash three times overfilling the wells with Wash buffer. After the final wash invert the plate and tap on absorbent filter papers or other absorbent material. It is recommended that the washer be set for a soaking time of 1 min.

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8. Add 100 µl of HRP Streptavidin Conjugate (SABC) working solution into each well, cover the plate with a new sealer and incubate at 37°C for 30 minutes.
9. Remove the cover and wash the plate 5 times with Wash buffer. Allow the wash buffer to remain in the wells 1-2 min for each wash. Discard the washing buffer and blot the plate onto absorbent filter papers or other absorbent material.
10. Add 90 µl of TMB substrate into each well. Cover the plate and incubate at 37°C in dark conditions for 15-20 minutes (incubation time is for reference only, do not exceed 30 minutes). When an apparent gradient appears in the standard wells the reaction can be terminated.
11. Add 50 µl of Stop solution into each well and mix thoroughly. The color should change to yellow immediately.
12. Read the O.D. absorbance at 450 nm in a microplate reader within 30' of adding the stop solution.

For calculation, (the relative O.D.₄₅₀) = (the O.D.₄₅₀ of each well) – (the O.D.₄₅₀ of Zero well). The standard curve can be plotted as the relative O.D.₄₅₀ of each standard solution (Y) vs. the respective concentration of the standard solution (X). The 5-HIAA concentration of the samples can be extrapolated from the standard curve.

Note: If the samples measured were diluted, multiply the dilution factor to the concentrations from extrapolation to obtain the concentration before dilution.

C. Precautions

1. Before using the kit, centrifuge the tubes briefly to bring down the contents trapped in the lid.
2. It is recommended to assay all standards, controls and sample in duplicate.
3. Do NOT let the plate dry out completely as this will inactivate the biological material on the plate.
4. To avoid cross contamination do not reuse pipette tips and tubes.
5. Do not use components from a different kit or expired ones.
6. The substrate is supplied as a ready to use solution. The TMB substrate is light sensitive and should be protected from direct sunlight and UV sources. Unreacted substrate should be colorless or very light yellow in appearance. Equilibrate to room temperature (25°C) prior to use, aspirate the dosage needed with sterilized tips and do not dump the residual solution back into the vial.

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D. Typical Data & Standard Curve

Typical Standard Curve Data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.

ng/ml	0	1.57	3.13	6.25	12.5	25	50	100
OD450	2.163	1.447	0.944	0.621	0.359	0.250	0.107	0.049

